Fecal microbiota transplantation induces remission of infantile allergic colitis through gut microbiota re-establishment

Liu, Sheng-Xuan; Li, Yin-Hu; Dai, Wen-Kui; Li, Xue-Song; Qiu, Chuang-Zhao; Ruan, Meng-Ling; Zou, Biao; Dong, Chen; Liu, Yan-Hong; He, Jia-Yi; Huang, Zhi-Hua; Shu, Sai-Nan

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ABSTRACTING

World Journal of Gastroenterology (WJG) is now indexed in Current Contents®/Clinical Medicine, Science Citation Index Expanded (also known as SciSearch®), Journal Citation Reports®, Index Medicus, MEDLINE, PubMed, PubMed Central and Directory of Open Access Journals. The 2017 edition of Journal Citation Reports® cites the 2016 impact factor for WJG as 3.365 (5-year impact factor: 3.176), ranking WJG as 29th among 79 journals in gastroenterology and hepatology (quartile in category Q2).

INDEXING

World Journal of Gastroenterology (WJG), print ISSN 1007-9327, online ISSN 2219-2840, DOI: 10.3748, is a peer-reviewed open access journal. WJG was established on October 1, 1995. It is published weekly on the 7th, 14th, 21st, and 28th each month. The WJG Editorial Board consists of 1375 experts in gastroenterology and hepatology from 68 countries.

The primary task of WJG is to rapidly publish high-quality original articles, reviews, and commentaries in the fields of gastroenterology, hepatology, gastrointestinal endoscopy, gastrointestinal surgery, hepatobiliary surgery, gastrointestinal oncology, gastrointestinal radiation oncology, gastrointestinal imaging, gastrointestinal interventional therapy, gastrointestinal infectious diseases, gastrointestinal pharmacology, gastrointestinal pathophysiology, gastrointestinal pathology, evidence-based medicine in gastroenterology, pancreatogastroenterology, gastrointestinal laboratory medicine, gastrointestinal molecular biology, gastrointestinal immunology, gastrointestinal microbiology, gastrointestinal genetics, gastrointestinal translational medicine, gastrointestinal diagnostics, and gastrointestinal therapeutics. WJG is dedicated to become an influential and prestigious journal in gastroenterology and hepatology, to promote the development of above disciplines, and to improve the diagnostic and therapeutic skill and expertise of clinicians.

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Retrospective Study

Fecal microbiota transplantation induces remission of infantile allergic colitis through gut microbiota re-establishment

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Author contributions: Liu SX and Li YH contributed equally to this work; Shu SN and Huang ZH designed the research; Dai WK designed the follow-up plans and collected patients’ information with Li XS, Ruan ML, Zou B and Dong C after FMT; Qiu CZ, Liu YH and He JY performed bioinformatics analysis; Ruan ML and Zou B recorded patients’ and donors’ clinical data; Shu SN and Huang ZH conducted FMT.

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Informed consent statement: All studied participants, or their legal guardians, provided informed written consent prior to study enrollment.

Conflict-of-interest statement: All authors declare that they have no competing interests.

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Abstract

AIM

To investigate the impact of fecal microbiota trans-
plantation (FMT) treatment on allergic colitis (AC) and gut microbiota (GM).

METHODS
We selected a total of 19 AC infants, who suffered from severe diarrhea/hematochezia, did not relieve completely after routine therapy or cannot adhere to the therapy, and were free from organ congenital malformations and other contraindications for FMT. Qualified donor-derived stools were collected and injected to the AC infants via a rectal tube. Clinical outcomes and follow-up observations were noted. Stools were collected from ten AC infants before and after FMT, and GM composition was assessed for infants and donors using 16S rDNA sequencing analysis.

RESULTS
After FMT treatment, AC symptoms in 17 infants were relieved within 2 d, and no relapse was observed in the next 15 mo. Clinical improvement was also detected in the other two AC infants who were lost to follow-up. During follow-up, one AC infant suffered from mild eczema and recovered shortly after hormone therapy. Based on the 16S rDNA analysis in ten AC infants, most of them (n = 6) had greater GM diversity after FMT. As a result, Proteobacteria decreased (n = 6) and Firmicutes increased (n = 10) in post-FMT AC infants. Moreover, Firmicutes accounted for the greatest proportion of GM in the patients. At the genus level, Bacteroides (n = 6), Escherichia (n = 8), and Lactobacillus (n = 4) were enriched in some AC infants after FMT treatment, but the relative abundances of Clostridium (n = 5), Veillonella (n = 7), Streptococcus (n = 6), and Klebsiella (n = 8) decreased dramatically.

CONCLUSION
FMT is a safe and effective method for treating pediatric patients with AC and restoring GM balance.

Key words: Pediatric; Infantile allergic colitis; Fecal microbiota transplantation; Gut microbiota; Immune reaction

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Core tip: This retrospective study explored the therapeutic effects and safety of fecal microbiota transplantation (FMT) treatment in 19 allergic colitis (AC) infants who were younger than 1 year old. After FMT treatment, AC symptoms were relieved in the patients rapidly, and no patient relapsed within 15 mo. With gut microbiota (GM) analysis, six of ten patients exhibited higher microbial diversity after FMT treatment. Moreover, decreased Proteobacteria and increased Firmicutes supplied the hints of GM re-establishment in the patients after FMT treatment. Therefore, this work showed the curative effects of FMT in AC infants and its possible mechanism.

INTRODUCTION
Allergic colitis (AC) is a common infantile rectal bleeding disorder which is caused by severe allergic reactions within the digestive system[1-2]. AC is normally identified in infants younger than one year of age and its representative clinical features are hematochezia and diarrhea[3]. Bloody purulent stools, abdominal pain, and vomiting are also used to diagnose AC[3]. Various factors such as food allergens, aberrant immune system, and imbalanced gut microbiota (GM) are thought to contribute to AC[4-6].

Conventional therapies for AC are reducing exposure to suspicious allergens and applying hypoallergenic milk powder[7]. Baldassarre et al[8] used probiotics to treat AC infants and found that Lactobacillus GG may relieve symptoms of AC by altering GM composition[8]. Fecal microbiota transplantation (FMT) can change gut micro-ecology more robustly in comparison to food or probiotics. Several reports suggested that FMT was therapeutically efficacious for treating diseases associated with GM dysbiosis, such as Clostridium difficile infection (CDI)[9,10], inflammatory bowel disease (IBD)[11,12], and irritable bowel syndrome (IBS)[13,14]. However, to our knowledge, FMT has not been used to treat AC infants.

Thus, we assessed 19 AC infants with severe hematochezia and/or diarrhea, who had not acquired complete remission after 2 wk of routine therapy or because the guardians cannot adhere to the routine therapy thoroughly. Our intention was to confirm the safety and efficacy of FMT in AC treatment, and detect the sustained GM changes after FMT.

MATERIALS AND METHODS

Ethics
This study was approved by the Medical Ethics Committee of Tongji Hospital, Tongji Medical College, Huazhong University of Science and Technology (TJ-C20140712). All subjects and donors gave signed informed consent. Principles of patients care and all experimental procedures followed the guidelines established by the Institutional Review Board of Tongji Hospital, Tongji Medical College, Huazhong University of Science and Technology.
Patient selection
AC was diagnosed based on the following clinical symptoms: (1) rectal bleeding with/without mucus and diarrhea; (2) exclusion of infectious colitis, anal fissure, lymphoid nodular hyperplasia, and uncommon conditions such as necrotizing enterocolitis, Hirschsprung’s enterocolitis, IBD, and IBS; (3) clinical remission after milk exclusion and recurrence after milk re-challenge; and (4) histological examination indicated that the intestinal mucosa exhibited chronic inflammation with eosinophils infiltration and colonic lesions (Supplementary File 1). Pediatric AC patients meeting the following criteria were selected as FMT candidates: (1) the patients had no complete remission after routine therapy, the patients cannot adhere to the therapy thoroughly, or their parents had strong intention to receive the treatment of FMT; (2) free from contraindications for FMT, such as intestinal obstructions, perforations, and bleeding, or severe immunodeficiency diseases; and (3) colonoscopic inspection indicated no mucosal congestion, edema, multiple spot-like erosion, or lymphoid granular nodes (four cases included in Figure 1). As a result, 19 AC patients were enrolled in the study between September 2015 and December 2015 (Table 1).

Donor screening
Patients’ mothers were considered to be donors of the highest priority, followed by fathers and healthy peers. Adult donors were screened as follows: (1) no infectious diseases history (e.g., tuberculosis and hepatopathy); (2) no metabolic diseases history (e.g., obesity and diabetes); (3) no gastrointestinal diseases (e.g., diarrhea, constipation, IBD, IBS, colorectal polyps, and gastrointestinal tumors); (4) no allergic diseases (e.g., food allergy, eczema, and allergic gastroenteritis); (5) no antibiotic exposure in the last 3 months; (6) no mental disorders or autoimmune diseases; and (7) no drug abuse history, amenorrhea (for mother donors), or psychological imbalance.

Candidate donors of the same age were selected with the following criteria: (1) preferred relatives with breast milk fed and same gender; (2) no antibiotic treatment in the last 3 months; (3) no allergic disease (e.g., food allergy, eczema, and allergic gastroenteritis); (4) no gastrointestinal disease (e.g., diarrhea, constipation, IBD, IBS, colorectal polyps, and gastrointestinal tumors); (5) no metabolic disease history (e.g., obesity and diabetes); and (6) no infectious disease history (e.g., tuberculosis, hepatopathy, and meases), and normal health and development. Tests for serum biochemistry and stool were performed for donors to ensure subject safety (Table 2).

FMT procedure
The application of parenteral nutrition and probiotics was stopped as soon as FMT began in the patients. No bowel preparation (cleanout or antibiotic pretreatment) was used prior to FMT, but pre-FMT clinical tests were performed as described in Table 3. Donor stool, collected 2 h before FMT, was diluted and mixed with sterile saline (1 mg of stool was diluted with 3 mL of saline). Samples were filtered through sterile gauze and 30-50 mL fecal suspension was prepared for FMT. FMT was administered over 5-10 min via a rectal tube into the left colon. The rectal tube was removed 15 min after administration and the fecal suspension was retained in the recipients’ gut for 4-6 h. Multi-FMT was given for patients with severe symptoms (Table 1).

Follow-up
Clinical symptoms, stool frequency, symptom remission time, and adverse events (e.g., abdominal pain, gastrointestinal infection, constipation, fever, and allergic disease) were recorded at the end of FMT (Table 1). Follow-up was conducted at ≥ 15 mo after FMT, except for two cases with 0.3 and 0.5 mo follow-up (AC17 and AC19), to evaluate FMT efficacy and safety (Table 1). The remission of AC was defined as the cease of rectal bleeding and decreased stool frequency (no more than two times/d) in the patients. The primary endpoint was the improved AC symptoms and sustained clinical remission at 12 mo. Secondary endpoint was the safety of FMT which was implied by the occurrence of adverse events.

Microbiota analysis and statistics
Fecal microbiota was analyzed for ten patients before FMT and during follow-ups. Donor feces were also assayed for GM. Microbial DNA was extracted using a PowerSoil DNA Isolation Kit (Mo Bio Laboratories, Carlsbad, CA) according to the manufacturer’s protocol and the hyper-variable V3-V4 region was amplified using 338F (5’-ACTCCTACGGGAGGCAGCA-3’) and 806R (5’-GGACTACHVGGGTWTCTAA-3’) primers. Library construction and sequencing were conducted on an Illumina MiSeq platform (Illumina, San Diego, United States). Data filtration and sequencing were performed as a prior report with the RDP database as an annotation reference. A Wilcoxon signed-rank test was used to compare samples of one patient, which were collected at different time points, and a Wilcoxon rank-sum test was used to compare donor and patient samples. Graphs were produced with R.
patients, and decreased defecation frequency with improved stool consistency was also observed (Table 1). Within more than 15 mo of follow-up, the symptoms of AC had not relapsed except two patients who were lost to follow-up (AC17 and AC19). Only one patient suffered from eczema, which appeared 2 mo after FMT and was resolved with hormone therapy. Beyond this, no other adverse event was recorded during FMT or the follow-up.

FMT treatment associated microbiota changes

Figures 2 and 3 depict microbiota changes of ten patients before and after FMT compared to donors. Microbiota diversity increased dramatically in five patients while it decreased in three patients after FMT (Figure 2). When sampled 1 or 2 mon after FMT, microbiota of six patients was more similar to donors’ microflora by comparison with pre-FMT samples (Figure 3).

After FMT treatment, Firmicutes accounted for the greatest proportion of GM in the AC infants, followed
Table 1  Clinical information for 19 allergic colitis infants

<table>
<thead>
<tr>
<th>ID</th>
<th>Gender</th>
<th>Age (mo)</th>
<th>Symptom(s)</th>
<th>Duration of disease (mo)</th>
<th>Treatment(s) before FMT</th>
<th>Donor source</th>
<th>FMT times</th>
<th>Symptom remission after first FMT (d)</th>
<th>Stool frequency before and after FMT (times/d)</th>
<th>Follow-up (mo)</th>
<th>Availability of gut microbiota data</th>
</tr>
</thead>
<tbody>
<tr>
<td>AC1</td>
<td>Female</td>
<td>7</td>
<td>Diarrhea, hematochezia sometimes</td>
<td>&gt; 3</td>
<td>Applying amino acid formula and probiotics (C. butyricum)</td>
<td>Mother</td>
<td>2</td>
<td>1</td>
<td>3-4, 1species/d</td>
<td>19</td>
<td>Yes</td>
</tr>
<tr>
<td>AC2</td>
<td>Male</td>
<td>10</td>
<td>Hematochezia</td>
<td>&gt; 0.5</td>
<td>Applying amino acid formula and probiotics (Bifidobacteria)</td>
<td>Healthy infants aged 10 mo old</td>
<td>2</td>
<td>1</td>
<td>2-3, 1species/d</td>
<td>18</td>
<td>Yes</td>
</tr>
<tr>
<td>AC3</td>
<td>Female</td>
<td>11</td>
<td>Hematochezia</td>
<td>&gt; 3</td>
<td>Applying amino acid formula and probiotics (S. boulardii)</td>
<td>Healthy infants aged 8 mo old</td>
<td>3</td>
<td>1</td>
<td>5-6, 2species/d</td>
<td>19</td>
<td>Yes</td>
</tr>
<tr>
<td>AC4</td>
<td>Male</td>
<td>9</td>
<td>Hematochezia</td>
<td>&gt; 3</td>
<td>Applying amino acid formula and probiotics (S. boulardii)</td>
<td>Mother's cousin sister</td>
<td>3</td>
<td>1</td>
<td>6-7, 1-2species/d</td>
<td>18</td>
<td>Yes</td>
</tr>
<tr>
<td>AC5</td>
<td>Male</td>
<td>5</td>
<td>Diarrhea and hematochezia sometimes</td>
<td>&gt; 3</td>
<td>Applying amino acid formula and probiotics (S. boulardii)</td>
<td>Healthy infants aged 8 mo old</td>
<td>3</td>
<td>1</td>
<td>3-4, 2species/d</td>
<td>19</td>
<td>Yes</td>
</tr>
<tr>
<td>AC6</td>
<td>Male</td>
<td>5</td>
<td>Hematochezia</td>
<td>&gt; 3</td>
<td>Applying amino acid formula</td>
<td>Mother</td>
<td>1</td>
<td>2</td>
<td>5-6, 1species/d</td>
<td>18</td>
<td>Yes</td>
</tr>
<tr>
<td>AC7</td>
<td>Male</td>
<td>4</td>
<td>Hematochezia and cough sometimes</td>
<td>&gt; 2</td>
<td>Applying amino acid formula and probiotics (S. boulardii)</td>
<td>Mother</td>
<td>2</td>
<td>2</td>
<td>4-7, 1species/d</td>
<td>15</td>
<td>Yes</td>
</tr>
<tr>
<td>AC8</td>
<td>Female</td>
<td>3</td>
<td>Diarrhea and mucoid feces sometimes</td>
<td>&gt; 2</td>
<td>Applying amino acid formula</td>
<td>Mother</td>
<td>2</td>
<td>1</td>
<td>3-4, 1-2species/d</td>
<td>19</td>
<td>Yes</td>
</tr>
<tr>
<td>AC9</td>
<td>Male</td>
<td>11</td>
<td>Interval hematochezia</td>
<td>&gt; 6</td>
<td>Applying amino acid formula and probiotics (S. boulardii)</td>
<td>Mother</td>
<td>2</td>
<td>1</td>
<td>3-4, 2species/d</td>
<td>23</td>
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<tr>
<td>AC10</td>
<td>Female</td>
<td>3</td>
<td>Hematochezia</td>
<td>&gt; 1.5</td>
<td>Applying amino acid formula</td>
<td>Healthy infants aged 10 mo old</td>
<td>4</td>
<td>2</td>
<td>5-6, 1species/d</td>
<td>21</td>
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<tr>
<td>AC11</td>
<td>Male</td>
<td>7</td>
<td>Diarrhea</td>
<td>&gt; 2</td>
<td>Applying amino acid formula, probiotics (Bifidobacteria), Simects and Oral Rehydration Salts (ORS)</td>
<td>Healthy infants aged 10 mo old</td>
<td>5</td>
<td>1</td>
<td>5-6, 1species/d</td>
<td>23</td>
<td>No</td>
</tr>
<tr>
<td>AC12</td>
<td>Female</td>
<td>10</td>
<td>Diarrhea and hematochezia sometimes</td>
<td>&gt; 1</td>
<td>Applying, amino acid formula and probiotics (Bifidobacteria)</td>
<td>Mother</td>
<td>3</td>
<td>1</td>
<td>5-6, 1-2species/d</td>
<td>22</td>
<td>No</td>
</tr>
<tr>
<td>AC13</td>
<td>Male</td>
<td>5</td>
<td>Hematochezia and diarrhea sometimes</td>
<td>&gt; 3</td>
<td>Applying amino acid formula</td>
<td>Mother</td>
<td>1</td>
<td>1</td>
<td>3-4, 1species/d</td>
<td>15</td>
<td>No</td>
</tr>
<tr>
<td>AC14</td>
<td>Female</td>
<td>5</td>
<td>Hematochezia and then peptone shaped feces</td>
<td>&gt; 1</td>
<td>Applying amino acid formula and probiotics (Bifidobacteria)</td>
<td>Mother</td>
<td>1</td>
<td>1</td>
<td>7-8, 2species/d</td>
<td>15</td>
<td>No</td>
</tr>
<tr>
<td>AC15</td>
<td>Male</td>
<td>7</td>
<td>Diarrhea</td>
<td>&gt; 2</td>
<td>Applying amino acid formula and probiotics (C. butyricum)</td>
<td>Mother</td>
<td>5</td>
<td>1</td>
<td>5-6, 1species/d</td>
<td>21</td>
<td>No</td>
</tr>
<tr>
<td>AC16</td>
<td>Female</td>
<td>5</td>
<td>Interval hematochezia</td>
<td>&gt; 2</td>
<td>Applying amino acid formula</td>
<td>Healthy infants aged 8 mo old</td>
<td>2</td>
<td>1</td>
<td>4-5, 1species/d</td>
<td>21</td>
<td>No</td>
</tr>
<tr>
<td>AC17</td>
<td>Male</td>
<td>7</td>
<td>Diarrhea and hematochezia sometimes</td>
<td>&gt; 3</td>
<td>Applying amino acid formula, probiotics (Bifidobacteria and C. butyricum)</td>
<td>Healthy infants aged 11 mo old</td>
<td>1</td>
<td>2</td>
<td>3-4, 1-2species/d</td>
<td>0.5</td>
<td>No</td>
</tr>
<tr>
<td>AC18</td>
<td>Female</td>
<td>8</td>
<td>Diarrhea and cough sometimes</td>
<td>&gt; 4</td>
<td>Applying amino acid formula and probiotics (Bifidobacteria and C. butyricum)</td>
<td>Healthy infants aged 8 mo old</td>
<td>2</td>
<td>1</td>
<td>3-4, 2species/d</td>
<td>17</td>
<td>No</td>
</tr>
<tr>
<td>AC19</td>
<td>Male</td>
<td>5</td>
<td>Interval diarrhea</td>
<td>&gt; 4</td>
<td>Applying amino acid formula</td>
<td>Mother</td>
<td>4</td>
<td>1</td>
<td>3-4, 1species/d</td>
<td>0.3</td>
<td>No</td>
</tr>
</tbody>
</table>

AC: Allergic colitis; FMT: Fecal microbiota transplantation.
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The rarefaction of samples

AC1

AC2

AC3

AC4

AC5

AC6

AC7

AC8
by Bacteroidetes and Proteobacteria. Proteobacteria decreased dramatically to < 10% for most patients except four patients (Supplementary File 2). Whilst, Firmicutes increased in all patients (Supplementary File 2).

The relative abundance of Escherichia significantly increased in eight AC infants (Supplementary File 3). For four patients, Lactobacillus was enriched after FMT, but for three subjects, it was absent even after FMT. Possible pathogens including Clostridium and Klebsiella generally decreased after FMT. Clostridium and Klebsiella decreased in five and eight AC infants respectively (Supplementary File 3). The relative abundance of Streptococcus was lowered in six patients. Whilst, Veillonella was found decreased in seven patients and its relative abundance was no more than

Figure 2  Shannon rarefaction curves of gut microbiota from ten allergic colitis infants and their donors. Each image represents one AC infant, and each curve represents one fecal sample from a patient or the corresponding donor. Sample ID has three parts: ‘R’ or ‘D’ indicates AC infants or donors, ‘pre’ or ‘post’ represents the stools collected before or after FMT, and fecal collection date. Microbiota diversity in six patients (AC1, AC4, AC5, AC7, AC8, and AC9) increased after FMT treatment. AC: Allergic colitis; FMT: Fecal microbiota transplantation.

Table 2  Laboratory testing on donors

<table>
<thead>
<tr>
<th>Blood testing</th>
</tr>
</thead>
<tbody>
<tr>
<td>TORCH examinations: Detections on toxoplasmosis IgG, toxoplasmosis IgM, rubella virus IgG, rubella virus IgM, cytomegalovirus IgG, cytomegalovirus IgM, herpes simplex virus 1/2 IgG, and herpes simplex virus 1/2 IgM.</td>
</tr>
<tr>
<td>Detection on parvovirus B19.</td>
</tr>
<tr>
<td>Epstein-Barr virus examinations: Detections on Epstein-Barr virus capsid antigen IgA, Epstein-Barr virus capsid antigen IgG, Epstein-Barr virus capsid antigen IgM, Epstein-Barr virus early antigen IgG, and Epstein-Barr virus nuclear antigen IgG.</td>
</tr>
<tr>
<td>Blood type examination.</td>
</tr>
<tr>
<td>Lymphocyte subpopulation examination.</td>
</tr>
<tr>
<td>Food allergen examination (sIgE).</td>
</tr>
<tr>
<td>Hepatic and renal function examinations: Glutamic-pyruvic transaminase, glutamic oxalacetic transaminase, total protein, albumin, globulin, prealbumin, total bilirubin, direct bilirubin, indirect bilirubin, alkaline phosphatase, gamma glutamyltranspeptidase, total cholesterol, triglycerides, high-density lipoprotein, low density lipoprotein, apolipoprotein A1, apolipoprotein B, lactic dehydrogenase, calcium, corrected calcium, phosphorus, magnesium, urea, creatinine, triosyppurine, bicarbonate, total bile acid, 5-nucleotidase, α-L-fucosidase, cholinesterase, cystatin C, and lipase and amylase.</td>
</tr>
<tr>
<td>Mycobacterium tuberculosis antibody examination (or the enzyme-linked immuno-spot assay test for tuberculosis).</td>
</tr>
<tr>
<td>Detection on hepatitis A-IgM.</td>
</tr>
<tr>
<td>Qualifications of C-reactive protein and erythrocyte sedimentation rate.</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Stool testing</th>
</tr>
</thead>
<tbody>
<tr>
<td>Fecal routine examinations: Detections on fecal color, character, red blood cells, white blood cells, occult blood, parasite eggs, protozoan, fat ball, rotavirus antigen, and fungus.</td>
</tr>
<tr>
<td>Bacterial culture tests: Detections on Vibrio cholera, Salmonella, Shigella, Aeromonas, Plesiomonas, and pathogenic Escherichia coli.</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Other testing</th>
</tr>
</thead>
<tbody>
<tr>
<td>Chest X-ray.</td>
</tr>
<tr>
<td>Abdominal ultrasound scan.</td>
</tr>
<tr>
<td>Electrocardiographic examination.</td>
</tr>
</tbody>
</table>
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8% after FMT. *Bifidobacterium* kept decreased in seven AC infants after FMT, and increased in two cases.

**DISCUSSION**

We chiefly considered the curative effects of FMT therapy in 19 AC infants and microbiota changes during treatment. Stools from both infant and adult donors suggested the same efficacy, and it was noted that all subjects had relieved symptoms of hematochezia and/or diarrhea in 2 d after the first FMT treatment. Due to the longer illness time or severe clinical symptoms, 15 patients experienced multi-FMT for the sustained clinical remission. And the multiple FMT in these patients gave us the idea that artificially modified microbiota for the specified patient might elevate the efficiency of FMT and attenuate transplantation times in the future. After being discharged from hospital, the patients were advised to take hypoallergenic milk powder instead of formula, and most patients had no relapse of colitis within more than 15 mo of follow-up. The recurrence of eczema

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**Table 3** Laboratory testing of the patients before fecal microbiota transplantation

<table>
<thead>
<tr>
<th>Blood testing</th>
</tr>
</thead>
<tbody>
<tr>
<td>Stool testing: Detections on fecal color, character, red blood cells, white blood cells, occult blood, parasite eggs, protozoan, fat ball, rotavirus antigen, and fungus. Bacterial culture tests: Detections on Vibrio cholera, Salmonella, Shigella, Aeromonas, Plesiomonas, and pathogenic Escherichia coli. Other testing: Enteroscopic examination. Abdominal ultrasound scan (intestinal adhesion). Electrocardiographic examination.</td>
</tr>
</tbody>
</table>

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**Figure 3** Microbiota similarity between allergic colitis infants and their donors. Values in red indicate low microbiota similarity between two samples. Blue represents high microbiota similarity. The microbiota compositions of patients (AC1, AC2, AC4, AC5, AC8, AC7, AC8, and AC10) were more similar to their donors' composition after FMT treatment. One patient (AC9) had more and then less microbiota similarity and AC3 did not change in this regard. AC: Allergic colitis.
in one infant might be caused by the inflammatory reactions which were triggered by discontinuous intake of hypoallergenic milk powder.

Fecal microbiota was analyzed in ten patients and their donors. We noted that the microbiota diversity increased in six patients after FMT. For three other subjects, GM diversity decreased after an initial increase while all the patients demonstrated clinical improvement. Individual-specific GM changes suggested the effect of donor’s GM complexity and patient’s gut micro-ecology imbalance. Khoruts et al.\(^\text{[10]}\) also suggested that bacteria can be eliminated due to nutrient competition, antimicrobial peptide suppression, and immune-mediated colonization resistance during GM re-establishment.

Proteobacteria, which contain opportunistic pathogens\(^\text{[12]}\), decreased in six AC infants, and their relative abundance was less than 10% after FMT. In contrast, Firmicutes increased in all AC infants. Previous work implied that Firmicutes decreased in patients with Cohn’s disease\(^\text{[23]}\), and the proportion of Firmicutes was negatively associated with gastrointestinal inflammation\(^\text{[24]}\).

After FMT, the relative abundances of Bacteroides and Lactobacillus increased. Prior reports showed that species in Bacteroides could secrete polysaccharide A, which promoted the number of T regulatory (Treg) cells\(^\text{[25,26]}\). Interleukin (IL)-10 produced by Treg cells also eliminated inflammatory reactions and protected against infectious pathogens\(^\text{[25,26]}\). Lactobacillus can secrete lactic acid, increase the proportion of Treg cells, and relieve symptoms of AC\(^\text{[16,27]}\). Generally, the relative abundance of opportunistic pathogens decreased, including Veillonella, Streptococcus, Clostridium, and Klebsiella. Prior reports suggested that the combination of Veillonella and Streptococcus had been found in various GM systems and can augment IL-8, IL-6, and tumor necrosis factor-α (TNF-α) responses, which were associated with inflammatory reactions\(^\text{[20]}\). Clostridium can cause diarrhea via enterotoxin secretion, and Klebsiella was positively associated with macrophage migration-inhibitory factor (MIF) and affected host immunity\(^\text{[29]}\). However, GM imbalance and post-FMT improvement need more analysis to understand the mechanisms underlying AC improvements.

This study pioneered the application of FMT in AC treatment and provided important reference to understand microbiota changes before and after FMT. Although the results favor the application of FMT for AC treatment, it is still important to clarify whether AC symptoms can be improved in our future studies with larger sample size. Also, we will explore the microbiota changes at the gene or functional level before and after FMT, to further the understanding of GM imbalance and re-configuration during FMT treatment of infantile AC.

**ARTICLE HIGHLIGHTS**

**Research background**

Allergic colitis (AC), which is characterized as hematochezia and severe diarrhea, is caused by an intense allergic reaction of the digestive system. Currently, first-line therapies for AC patients are reducing exposure to suspicious allergens and applying hypoallergenic milk powder. However, some pediatric patients could not relieve from AC symptoms completely with routine treatment, and long-term illness causes adverse impact on nutrition absorption and physical development in the children.

**Research motivation**

Previous studies indicated that gut microbiota (GM) was closely related with the digestive system, neural system, and immune system in human. Meanwhile, the positive effects of fecal microbiota transplantation (FMT) have been confirmed in various gastrointestinal diseases, including Clostridium difficile infection (CDI), inflammatory bowel disease (IBD), and irritable bowel syndrome (IBS). However, FMT had not been applied to treat AC infants before. This research could provide important references for the treatment and research of infantile AC with FMT therapy.

**Research objectives**

The research aimed to detect the safety and efficiency of FMT treatment in AC, and compare GM composition before and after FMT treatment in the patients.

**Research methods**

The procedures of FMT, including selection of AC patients and donors, were conducted according to the guidelines established by the Institutional Review Board of Tongji Hospital, Tongji Medical College, Huazhong University of Science and Technology. Wilcoxon tests were adopted in the research.

**Research results**

In this study, the safety and efficacy of FMT treatment were investigated in 19 AC infants with GM analysis. The results indicated that the AC symptoms, which included rectal bleeding, diarrhea and hematochezia, were relieved rapidly by FMT treatment. During the 15 mo follow-up, no relapse was recorded except that eczema happened in one patient. After FMT treatment, the elevation of microbial diversity was detected in six of ten patients. Meanwhile, the relative abundances of Proteobacteria and Firmicutes were decreased (6/10) and increased (10/10), respectively, in the AC infants.

**Research conclusions**

This study documents the positive effect of FMT treatment on infantile AC remission, suggesting the potential of FMT in gastrointestinal allergic diseases. Individual-specific GM re-configuration also extended our understanding of FMT efficacy and associated mechanisms.

**Research perspectives**

Despite the aspiring results of FMT in pediatric AC, verified improvements with larger cohorts and longer follow-up are necessary. In parallel, GM analysis should be performed before and after FMT, to unravel keystones of microbial components in the specific disease.

**ACKNOWLEDGEMENTS**

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