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Complete Genome Sequences of Four *Campylobacter jejuni* Strains Isolated from Retail Chicken Meat and Broiler Feces

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ABSTRACT *Campylobacter jejuni* is the leading pathogen that causes foodborne infections. Here, we report the complete genome sequences of four *C. jejuni* strains isolated from retail chicken meat and broiler feces samples. Genes encoding type VI secretion and antibiotic resistance were detected among these isolates.

Campylobacteriosis, caused by *Campylobacter jejuni*, is one of the major foodborne infections that cause human gastroenteritis, and poultry is considered the major reservoir host (1–6). Therefore, to understand the variation in molecular characteristics of *C. jejuni* poultry isolates, whole-genome sequencing was performed to determine detailed information about the virulence and antimicrobial resistance genes (6–8).

Four *C. jejuni* strains isolated and identified in our previous study (9) were further utilized. Each *C. jejuni* strain was cultured in Bolton broth (Thermo Fisher Scientific, USA) and incubated under microaerophilic conditions for 48 h at 42°C; genomic DNA was then extracted using the GeneJET genomic DNA purification kit (Thermo Fisher Scientific). For long-read sequencing, genomic DNA was fragmented using a g-TUBE device (Covaris, USA) and the fragments size selected to a mean of 8 to 12 kb using AMPure XP beads (Beckman Coulter, USA) for library preparation. A multiplexing library pool was prepared and barcoded using a genomic DNA ligation kit (SQK-LSK109) and the native barcoding genomic DNA kit (SQK-NBD104), respectively; the library was sequenced on an R9.4 MinION flow cell using the Nanopore GridION sequencer (Oxford Nanopore Technologies, Oxford, UK) for 48 h. For short-read sequencing, Illumina TruSeq DNA PCR-free (insert size, 350 bp) library preparation and sequencing using the paired-end 150-bp (PE150) method was performed by Novogene on the HiSeq X Ten sequencer (Illumina, USA). The raw Nanopore reads were assembled into contigs using Canu v1.9 (10). The first 1,000 bp of each contig was mapped to the entire contig using blastn from the BLAST+ v2.9.0 tool suite (11); then, the overlapping regions were removed using SAMtools v1.9 (12). The start of each circular contig that had an overlapping region was moved upstream of the *dnaA* gene, when present, or upstream of the gene closest to the middle of the contig using Circlator v1.5.5 (13). The junction site of circular contigs was verified using PCR amplification and Sanger sequencing (14). The Illumina and Nanopore reads were mapped to the assembled contigs using BWA v0.7.17-r1188 (15) and minimap2 v2.17 (16), respectively. Any contig without supporting Illumina data was removed. The remaining contigs were polished using Pilon v1.23 (17). The genome sequences were annotated using Prokka v1.13 (18).

The genome sizes and number of protein-coding DNA sequences of the isolates range from 1.63 Mb to 1.84 Mb and 1,421 to 1,552, respectively, and two isolates

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TABLE 1 Summary of genome assembly and genotype characteristics of four *C. jejuni* isolates

Strain	Source	GenBank accession no.	Size (bp)	Illumina		Nanopore		GC content (%)	No. of CDSs ^a	ARG(s) ^b	Virulence genes	
				No. of reads	Avg per-base coverage (×)	No. of reads	Avg per-base coverage (×)					
<i>C. jejuni</i> MS2005 ^c	Retail chicken meat	CP084080	1,757,242	3,476,551	571	164,000	736	10,111	30.5	1,541	<i>aph(3')-IIIa</i> , <i>sat-4</i> , <i>cmeR</i>	<i>tssA</i> , <i>tssB</i> , <i>tssC</i> , <i>tssD</i> , <i>tssE</i> , <i>tssF</i> , <i>tssG</i> , <i>tssI</i> , <i>tssJ</i> , <i>tssK</i> , <i>tssL</i> , <i>tssM</i>
<i>C. jejuni</i> MS2005 ^d	Retail chicken meat	CP084081	5,208		6,746		405		28.5	6		
<i>C. jejuni</i> MS2058 ^c	Retail chicken meat	CP084082	1,758,823	5,455,985	890	104,000	472	10,256	30.5	1,552	<i>aph(3')-IIIa</i> , <i>sat-4</i> , <i>cmeR</i>	<i>tssA</i> , <i>tssB</i> , <i>tssC</i> , <i>tssD</i> , <i>tssE</i> , <i>tssF</i> , <i>tssG</i> , <i>tssJ</i> , <i>tssK</i> , <i>tssL</i> , <i>tssM</i>
<i>C. jejuni</i> MS2058 ^d	Retail chicken meat	CP084083	40,825		1,086		231		28.5	41	<i>tet(O)</i>	
<i>C. jejuni</i> MS2074 ^c	Retail chicken meat	CP084084	1,629,343	5,376,179	986	144,000	691	9,909	30.7	1,491	<i>bla</i> _{OXA-61} , <i>cmeR</i>	
<i>C. jejuni</i> MS2167 ^c	Broiler feces	CP084085	1,711,301	5,762,419	1,007	220,000	951	9,302	30.5	1,491	<i>cmeR</i>	<i>tssA</i> , <i>tssB</i> , <i>tssC</i> , <i>tssD</i> , <i>tssE</i> , <i>tssF</i> , <i>tssG</i> , <i>tssI</i> , <i>tssJ</i> , <i>tssK</i> , <i>tssL</i> , <i>tssM</i>

^a CDSs, protein-coding DNA sequences.

^b ARG(s), antimicrobial resistance gene(s).

^c Chromosome.

^d Plasmid.

(MS2005 and MS2058) contain one plasmid each. The respective fold coverages for Illumina and Nanopore ranged from 541 to 864 and 440 to 851, and the N_{50} values ranged from 9,302 to 10,256 bp, respectively. *In silico* prediction of antimicrobial resistance genes using CARD v3.1.0 (19) identified *aph(3')-IIIa*, *sat-4*, and *bla*_{OXA-61} genes in the chromosomes, whereas plasmid pMS2058 contains the *tet(O)* gene. Using SecReT6 (20), the presence of type VI secretion genes was predicted in the chromosomes of three *C. jejuni* strains MS2005, MS2058, and MS2167 (Table 1). The genome information obtained in this study can be further utilized for the identification of potential vaccine candidates and functional analysis of type VI secretion genes.

Data availability. The genome sequences and raw data are available at NCBI GenBank under the BioProject accession number PRJNA655459. The specific parameters and code used for sequencing can be found at https://github.com/IGBB/campylobacter_jejuni.

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REFERENCES

- Man SM. 2011. The clinical importance of emerging *Campylobacter* species. *Nat Rev Gastroenterol Hepatol* 8:669–685. <https://doi.org/10.1038/nrgastro.2011.191>.
- Kaakoush NO, Castaño-Rodríguez N, Mitchell HM, Man SM. 2015. Global epidemiology of *Campylobacter* infection. *Clin Microbiol Rev* 28:687–720. <https://doi.org/10.1128/CMR.00006-15>.
- Tack DM, Ray L, Griffin PM, Cieslak PR, Dunn J, Rissman T, Jervis R, Lathrop S, Muse A, Duwell M, Smith K, Tobin-D'Angelo M, Vugia DJ, Zablotsky Kufel J, Wolpert BJ, Tauxe R, Payne DC. 2020. Preliminary incidence and trends of infections with pathogens transmitted commonly through food—Foodborne Diseases Active Surveillance Network, 10 US Sites, 2016–2019. *MMWR Morb Mortal Wkly Rep* 69:509–514. <https://doi.org/10.15585/mmwr.mm6917a1>.
- Wieczorek K, Denis E, Osek J. 2015. Comparative analysis of antimicrobial resistance and genetic diversity of *Campylobacter* from broilers slaughtered in Poland. *Int J Food Microbiol* 210:24–32. <https://doi.org/10.1016/j.ijfoodmicro.2015.06.006>.
- Wilson DJ, Gabriel E, Leatherbarrow AJH, Cheesbrough J, Gee S, Bolton E, Fox A, Fearnhead P, Hart CA, Diggle PJ. 2008. Tracing the source of *Campylobacter* infection. *PLoS Genet* 4:e1000203. <https://doi.org/10.1371/journal.pgen.1000203>.
- Hale CR, Scallan E, Cronquist AB, Dunn J, Smith K, Robinson T, Lathrop S, Tobin-D'Angelo M, Clogher P. 2012. Estimates of enteric illness attributable to contact with animals and their environments in the United States. *Clin Infect Dis* 54:S472–S479. <https://doi.org/10.1093/cid/cis051>.
- Redondo N, Carroll A, McNamara E. 2019. Molecular characterization of *Campylobacter* causing human clinical infection using whole-genome sequencing: virulence, antimicrobial resistance and phylogeny in Ireland. *PLoS One* 14:e0219088. <https://doi.org/10.1371/journal.pone.0219088>.
- FDA. 2016. Whole genome sequencing (WGS) program. <https://www.fda.gov/food/science-research-food/whole-genome-sequencing-wgs-program>. Accessed 27 April, 2022.
- Poudel S, Li T, Chen S, Zhang X, Cheng W-H, Sukumaran AT, Kiess AS, Zhang L. 2022. Prevalence, antimicrobial resistance, and molecular

- characterization of *Campylobacter* isolated from broilers and broiler meat raised without antibiotics. *Microbiol Spectr* 10:e00251-22. <https://doi.org/10.1128/spectrum.00251-22>.
10. Koren S, Walenz BP, Berlin K, Miller JR, Bergman NH, Phillippy AM. 2017. Canu: scalable and accurate long-read assembly via adaptive k-mer weighting and repeat separation. *Genome Res* 27:722–736. <https://doi.org/10.1101/gr.215087.116>.
 11. Camacho C, Coulouris G, Avagyan V, Ma N, Papadopoulos J, Bealer K, Madden TL. 2009. BLAST+: architecture and applications. *BMC Bioinform* 10:421. <https://doi.org/10.1186/1471-2105-10-421>.
 12. Li H, Handsaker B, Wysoker A, Fennell T, Ruan J, Homer N, Marth G, Abecasis G, Durbin R, 1000 Genome Project Data Processing Subgroup. 2009. The Sequence Alignment/Map format and SAMtools. *Bioinformatics* 25:2078–2079. <https://doi.org/10.1093/bioinformatics/btp352>.
 13. Hunt M, De Silva N, Otto TD, Parkhill J, Keane JA, Harris SR. 2015. Circlator: automated circularization of genome assemblies using long sequencing reads. *Genome Biol* 16:294. <https://doi.org/10.1186/s13059-015-0849-0>.
 14. Li T, Castañeda CD, Arick MA, Hsu CY, Kiess AS, Zhang L. 2020. Complete genome sequence of multidrug-resistant avian pathogenic *Escherichia coli* strain APEC-O2-MS1170. *J Glob Antimicrob Resist* 23:401–403. <https://doi.org/10.1016/j.jgar.2020.11.009>.
 15. Li H. 2013. Aligning sequence reads, clone sequences and assembly contigs with BWA-MEM. arXiv 1303.3997v2 [q-bio.GN]. <https://doi.org/10.48550/arXiv.1303.3997>.
 16. Li H. 2018. Minimap2: pairwise alignment for nucleotide sequences. *Bioinformatics* 34:3094–3100. <https://doi.org/10.1093/bioinformatics/bty191>.
 17. Walker BJ, Abeel T, Shea T, Priest M, Abouelliel A, Sakthikumar S, Cuomo CA, Zeng Q, Wortman J, Young SK, Earl AM. 2014. Pilon: an integrated tool for comprehensive microbial variant detection and genome assembly improvement. *PLoS One* 9:e112963. <https://doi.org/10.1371/journal.pone.0112963>.
 18. Seemann T. 2014. Prokka: rapid prokaryotic genome annotation. *Bioinformatics* 30:2068–2069. <https://doi.org/10.1093/bioinformatics/btu153>.
 19. Alcock BP, Raphenya AR, Lau TTY, Tsang KK, Boucharde M, Adalatmand A, Huynh W, Nguyen A-LV, Cheng AA, Liu S, Min SY, Miroshnichenko A, Tran H-K, Werfalli RE, Nasir JA, Oloni M, Speicher DJ, Florescu A, Singh B, Faltyn M, Hernandez-Koutoucheva A, Sharma AN, Bordeleau E, Pawlowski AC, Zubyk HL, Dooley D, Griffiths E, Maguire F, Winsor GL, Beiko RG, Brinkman FSL, Hsiao WWL, Domselaar GV, McArthur AG. 2020. CARD 2020: antibiotic resistance surveillance with the Comprehensive Antibiotic Resistance Database. *Nucleic Acids Res* 48:D517–D525. <https://doi.org/10.1093/nar/gkz935>.
 20. Li J, Yao Y, Xu HH, Hao L, Deng Z, Rajakumar K, Ou HY. 2015. SecReT6: a Web-based resource for type VI secretion systems found in bacteria. *Environ Microbiol* 17:2196–2202. <https://doi.org/10.1111/1462-2920.12794>.